

Effect of Vitamin E on Serum Levels of Vascular Endothelial Growth Factor and Angiopoietin-1 in Women with Polycystic Ovary Syndrome: A Pilot Randomized, Placebo-Controlled Trial

Shabnam Shirazi, M.Sc.^{1,2}, Bahram Pourghassem Gargari, Ph.D.^{3*}, Azimeh Izadi, Ph.D.^{1,2}, Shiva Taghizadeh, M.Sc.^{1,2}, Marziyeh Parizad, M.D.⁴

1. Department of Biochemistry and Diet Therapy, Faculty of Nutrition and Food Sciences, Tabriz University of Medical Sciences, Tabriz, Iran

2. Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran

3. Nutrition Research Center, Department of Biochemistry and Diet Therapy, Faculty of Nutrition and Food Sciences, Tabriz University of Medical Sciences, Tabriz, Iran

4. Women's Reproductive Health Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

Abstract

Background: Angiogenesis disturbances are common in women with polycystic ovary syndrome (PCOS). Vitamin E has antiangiogenic properties. Data on the effects of vitamin E on angiogenesis in PCOS is limited, so the current study was conducted to evaluate its effects on angiogenic indices in PCOS patients.

Materials and Methods: This randomized, double-blind, placebo-controlled trial was performed on 43 women aged 20-40 years, diagnosed with PCOS (Rotterdam criteria). It was performed at the referral clinic affiliated to Tabriz University of Medical Sciences, Tabriz, Iran, from April 2017 to September 2017. Patients were randomly assigned into two groups to receive either 400 IU/day vitamin E -as alpha tocopheryl acetate- (n=22) or placebo (n=21), for 8 weeks. Anthropometric, and angiogenic parameters including body weight, fat mass and fat free mass, vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF), angiopoietin-1 (Ang-1), and angiopoietin-2 (Ang-2) were measured by standard methods at the beginning and at the end of study. Statistical Package for Social Science version 25 was used for statistical analysis and P<0.05 were considered significant.

Results: After adjusting for potential confounders, we observed that vitamin E supplementation significantly reduced body weight, fat mass, Ang-1, Ang-1/Ang-2 ratio and VEGF (P<0.01). We did not observe any considerable effect for vitamin E on Ang-2 level or bFGF.

Conclusion: Vitamin E supplementation for 8 weeks in the PCOS women had beneficial effects on body weight, Ang-1, Ang-1/Ang-2 ratio, and VEGF level (Registration number: IRCT201610193140N18).

Keywords: Angiopoietins, Basic Fibroblast Growth Factor, Polycystic Ovary Syndrome, Vascular Endothelial Growth Factor, Vitamin E

Citation: Shirazi Sh, Pourghassem Gargari B, Izadi A, Taghizadeh Sh, Parizad M. Effect of vitamin E on serum levels of vascular endothelial growth factor and angiopoietin-1 in women with polycystic ovary syndrome: A pilot randomized, placebo-controlled trial. *Int J Fertil Steril.* 2021; 15(1): 44-50. doi: 10.22074/IJFS.2020.45677.

This open-access article has been published under the terms of the Creative Commons Attribution Non-Commercial 3.0 (CC BY-NC 3.0).

Introduction

Polycystic ovary syndrome (PCOS) is one of the most complex endocrine disorders that causes infertility due to ovulation failure in women (1). Approximately 6-25% of women of reproductive age, are influenced by PCOS (2). The prevalence of PCOS in Iranian women was reported as 19.5% based on

the Rotterdam criteria (3). The clinical symptoms of PCOS include menstrual dysfunction, hyperandrogenism, polycystic ovaries, and subfertility (2). Additionally, PCOS can cause obesity and metabolic disorders such as insulin resistance, dyslipidemia, raised levels of inflammatory factors, and endothelial dysfunction. Long-term consequences

Received: 3 November 2019, Accepted: 11 August 2020

*Corresponding Address: P.O.Box: 5166614711, Nutrition Research Center, Department of Biochemistry and Diet Therapy, Faculty of Nutrition and Food Sciences, Tabriz University of Medical Sciences, Tabriz, Iran
Email: pourghassemb@tbzmed.ac.ir



Royan Institute
International Journal of Fertility and Sterility
Vol 15, No 1, January-March 2021, Pages: 44-50

of PCOS are endometrial cancer, diabetes mellitus, hypertension, and cardiovascular disorders (4, 5). The etiology of PCOS remains largely unknown, however, there is accumulating evidence suggesting that angiogenesis dysregulation might play the main role in the pathogenesis of PCOS (4). Angiogenesis is a complex physiological process where new vessels develop from preexisting vasculature (5). Angiogenesis in the ovary is an important part of the process of the menstrual cycle (6). An essential role of the formation of the new vessels in the ovary, is the provision of nutrients and hormones for development of the corpus luteum and follicular growth (4).

Vascular endothelial growth factors (VEGFs) and angiopoietins are among the most important angiogenic markers. Other indices include basic fibroblast growth factor (bFGF) -also known as fibroblast growth factor-2 (FGF2)- and platelet-derived growth factor (PDGF). Angiopoietin-1 and -2 (Ang-1 and Ang-2, respectively) as well as VEGF play major roles in the regulation of angiogenesis in the ovary (4).

It was suggested that PCOS women have imbalances in angiogenic/antiangiogenic indices with partial dominance of pro-angiogenic markers. In this regard, increased ovarian expression of VEGF and bFGF has been reported in PCOS women. In addition, elevated levels of Ang-1 were shown in PCOS women compared to the healthy controls (4, 7). The abnormal alterations can cause cysts in the ovary, and disrupt and reduce ovulation rates. The recovery of proper blood vessel development in the ovaries, could improve follicular growth as well as development and ovulation among patients with PCOS (8).

Several studies have suggested that tocopherols reduce the processes of inflammation and angiogenesis (9). In addition, vitamin E levels in the blood of women with PCOS were lower than those of healthy subjects (10). Rahmani et al. (11) reported that vitamin E co-supplementation with omega-3 fatty acids, significantly regulated lipid profile and reduced oxidative stress products in PCOS women. Vitamin E and D co-supplementation was shown to improve pregnancy outcome in PCOS women (12). In another study, vitamin E supplementation inhibited VEGF-A-mediated angiogenesis (13). In addition to the role of vitamin E in angiogenesis, some evidence indicated that vitamin E has an association with obesity (14). It seems that the mentioned effects are not due to the antioxidant mechanism of vitamin E (12).

Considering data scarcity in this subject, the present study was conducted to evaluate the effect of vitamin E on serum VEGF, bFGF, Ang-1, and Ang -2 as well as Ang-1/Ang-2 ratio in PCOS women. We hypothesized that vitamin E supplementation might have an effect on the angiogenic markers and imbalances in patients with PCOS.

Materials and Methods

This double-blinded, placebo-controlled clinical trial was part of a larger study approved by the Ethics Committee of Tabriz University of Medical Sciences (Ethics approval No. IR.TBZMED.REC.1395.777) and registered at Iranian Registry of IRCT (IRCT201610193140N18). It was performed at the referral clinic affiliated to the Tabriz University of Medical Sciences, Tabriz, Iran from April 2017 to September 2017. The study was advertised in different clinical and therapeutic centers. For the present study, the sample size was calculated based on the results of blood VEGF concentration reported by Mondul et al. (13) by using G*Power (version 3.1.2, Germany). The number of participants was calculated as at least 16 subjects in each group. Considering dropout and to ensure a sample size sufficiently large to enable reliable estimates, we enrolled 22 and 21 subjects in vitamin E and placebo groups, respectively.

The volunteers were given more details on the study by the first author, and then, a written consent form was signed by all of the participants. The participants were able to withdraw from the study at any time.

The inclusion criteria of the study were women within the age range 20-40 years who were diagnosed with PCOS in accordance with the Rotterdam criteria (15). On the other hand, menopause, pregnancy or lactation, diabetes, having hepatic, renal, thyroid, coagulation or cardiovascular disorders, elevated levels of prolactin, smoking, alcohol consumption, fat malabsorption, receiving oral anticoagulants, ovulation induction agents or drugs affecting hormonal profile such as oral contraceptive pills (OCP), or having taken antioxidant supplements or adopted a diet or a particular plan for physical activity within the last 3 months, were considered exclusion criteria (16).

Trial design

Trial design was parallel. Initially, forty-three PCOS women with $25 \leq$ body mass index (BMI) < 35 kg/m², enrolled in to the study. The participants were randomly assigned into one of the two groups (in a 1:1 ratio), using the Random Allocation Software. The subjects in vitamin E and placebo groups received 400 IU/day vitamin E -as alpha tocopheryl acetate- (n=22), or cellulose capsules (n=21), for 8 weeks.

Vitamin E capsules were produced by Nature Made Pharmaceutical Company (USA, Batch number: 1143156) and provided by Pourateb Pharmaceutical Company (Iran). The placebo capsules were made by Barij Essence Pharmaceutical Corporation (Iran). The capsules of vitamin E and placebo were similar in size and shape. The patients and researcher were blind to allocations until the end of the study. Based on the guidelines (17), all patients received metformin at the dose of 1500 mg (500 mg 3 times daily). At the baseline of the study, the patients were asked to keep their physi-

cal activity and diet unchanged within the 8 weeks of intervention.

Adherence to the study

To assess the compliance, the participants were requested to bring the medication containers. All patients were monitored by a weekly phone call and encouraged to consume the supplement. Short Message Service was sent to the patients' cell phones every day. To check the adherence to treatments, the participants were asked to bring the unused capsules. The subjects who had incomplete consumption of the drugs (less than 90% consumption) were excluded from the study.

Evaluation of anthropometrics

Body weight (following overnight fasting) was measured by a digital scale (Seca, Hamburg, Germany) with an accuracy of ± 0.1 kg. Height was measured by a non-elastic strip (Seca, Germany) with a precision of 0.1 cm. Further, BMI was calculated as weight in kilograms divided by squared height in meters. The body composition indices including body fat mass percentage (FM%), fat mass (FM), and fat free mass (FFM), were evaluated by a bioelectrical impedance analyzer (8-electrode, TanitaBC-418 MA; Tanita Co., Japan).

Assessment of dietary intake and physical activity

Dietary intake was assessed by 24-hour recall, which was completed on three different days of the week (two weekdays and one weekend). To assess the nutrient intake of the patients, Nutritionist IV software (First Databank, CA) edited for Iranian foods, was used. To control the confounding effects of physical activity, international physical activity questionnaire-short form- (IPAQ-S) was employed for evaluation of physical activity (18). The validity and reliability of the Persian translation of IPAQ in previous studies on the Iranian populations, were tested and approved (19). We assessed physical activity and dietary intakes at the baseline and at the 8th week of intervention.

Laboratory analysis

At the beginning and the end of the study, the patients were instructed to refer to the laboratory on days 3 to 5 of normal menstrual cycle or menstrual induced by progesterone. Fasting blood samples were obtained from the participants. To separate the serum, centrifugation at 1200 rpm for 12 minutes, was done. The samples were kept at -80°C for subsequent experiments (20). Serum levels of Ang-1 and Ang-2, VEGF, and bFGF were measured using the commercial Enzyme-Linked Immunosorbent Assays (ELISAs, Bioassay Technology Laboratory, Shang-

hai Korain Biotech, China) according to the manufacturer's instructions. Coefficients of variation of the intra-assay and inter-assay assays were less than 10%.

Statistical methods

Data analysis was performed by an intention-to-treat procedure where missing values were treated based on Last-Observation-Carried-Forward method. To identify within-group differences (pre- and post-intervention), we utilized paired samples t tests. To determine between group differences, independent sample t test and Mann-Whitney U test were used for comparison of normally and abnormally distributed variables, respectively. To identify the impacts of vitamin E supplementation on anthropometric and biochemical variables, analysis of covariance adjusted for age, physical activity, BMI, and baseline values, was employed. We used Statistical Package for Social Science version 25 (SPSS Inc., Chicago, Illinois, USA) for all statistical analyses, with $P < 0.05$ considered significant.

Results

Four subjects dropped out the study because of pregnancy ($n=1$) or in vitro fertilization ($n=3$). Finally, 39 participants remained in the study. However, intention-to-treat analysis was used, so, data for 43 PCOS women were included in the final analysis (Fig.1). No major side effect was observed following taking vitamin E supplement. Hyperandrogenism (clinically) and PCOS (by ultrasound) were seen in nearly all of the subjects. Ninety percent of the participants had oligo-anovulation. The compliance rate of the studied groups was more than 90%. The baseline characteristics of the participants are shown in Table 1. There were no significant differences between the two groups in terms of age, height, and physical activity. Baseline measures for follicle-stimulating hormone (FSH), and luteinizing hormone (LH) were not different between the vitamin E and placebo groups. More details on the hormonal status of the subjects are given in another published article (16).

Table 1: Baseline characteristics of the studied subjects in the vitamin E and placebo groups

Variables	Vitamin E n=22	Placebo n=21	P value*
Age (Y)	27.18 \pm 5.77	26.0 \pm 4.53	0.68
Height (cm)	162.27 \pm 6.86	159.81 \pm 6.06	0.22
Physical activity (MET-minute/week)			
Low	8 (36.4)	10 (47.6)	
Moderate	11 (50)	7(52.4)	0.54**
Vigorous	3(13.6)	4 (19)	

Data are presented as mean \pm SD or n (%).*; Assessed by independent t test and **; Chi-square test, and METs; Metabolic equivalents.

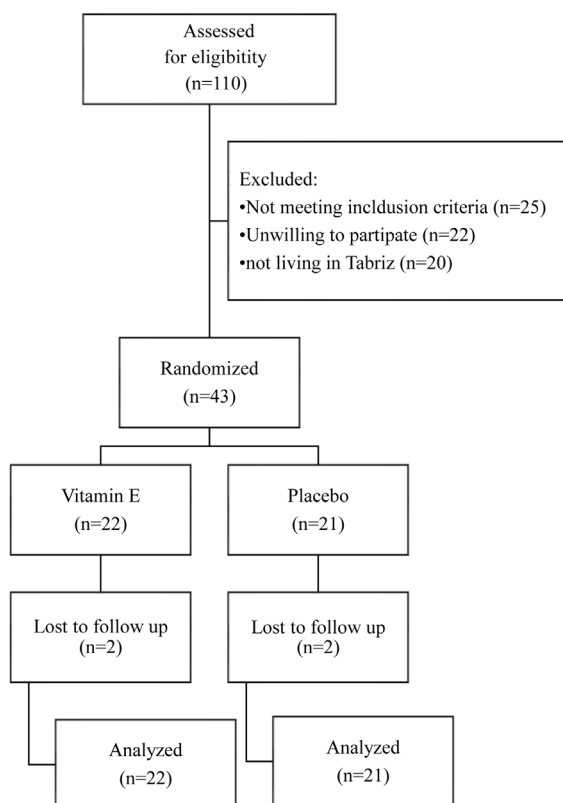


Fig.1: Flowchart of the study.

Dietary intake

Table 2 presents dietary intakes of the studied subjects. There were no significant differences in the dietary intakes of energy and nutrients between the two studied groups.

Table 2: Dietary intakes of the study participants throughout the study in the vitamin E and placebo groups

Variables	Vitamin E n=22	Placebo n=21	P value*
Energy (Kcal/day)	1698.46 ± 215.88	1745.87 ± 308.00	0.56
Carbohydrate (g/day)	214.89 ± 28.03	227.63 ± 52.57	0.32
Protein (g/day)	66.65 ± 13.66	56.9 ± 12.77	0.36
Fat (g/day)	68.05 ± 15.75	63.57 ± 18.47	0.39
SFAs (g/day)	16.09 ± 7.71	15.28 ± 7.48	0.73
PUFAs (g/day)	13.37 ± 5.89	12.69 ± 6.66	0.72
MUFAs (g/day)	17.2 ± 6.3	16.94 ± 6.59	0.89
Cholesterol (mg/day)	194.98 ± 58.55	206.21 ± 65.66	0.56
Fiber (g/day)	20.73 ± 4.79	20.31 ± 4.71	0.77
Vitamin E (mg/day)	6.09 ± 3.1**	6.85 ± 3.23	0.47
Vitamin A (RE/d)	440.12 ± 81.93	422.65 ± 132.51	0.60
Vitamin C (mg/d)	66.45 ± 12.05	73.80 ± 20.09	0.15
Selenium (µg/d)	43.39 ± 4.26	48.29 ± 20.80	0.30
Zinc (mg/day)	5.23 ± 1.34	5.21 ± 0.99	0.94

Data are presented as mean ± SD. SFA; Saturated fatty acid, PUFA; Polyunsaturated fatty acid, MUFA; Monounsaturated fatty acid, *; Assessed by independent t test, and **; Vitamin E level is estimated based only on dietary consumption, in the absence of the study supplement.

Anthropometric measurements

No significant difference was found at the baseline of the study in the assessed anthropometric indices except for FM which was significantly higher in the vitamin E group. In within-group analysis, all assessed anthropometric indices had significant changes in the vitamin E supplemented group ($P < 0.01$). In between-groups comparisons, except for FFM, the assessed anthropometric indices were reduced in the vitamin E-supplemented group compared to the placebo group (Table 3).

Table 3: Baseline and 8 weeks after intervention values of the anthropometric indices in the vitamin E and placebo groups

Variables	Vitamin E n=22	Placebo n=21	P value
Weight (kg)			
Before	76.95 ± 10.61	73.23 ± 7.58	0.19 ^b
After	75.96 ± 10.3	73.29 ± 7.3	0.01 ^c
P value ^a	0.003	0.82	
BMI (kg/m ²)			
Before	29.45 ± 5.35	28.80 ± 3.71	0.64 ^b
After	29.07 ± 5.16	28.83 ± 3.70	0.01 ^c
P value ^a	0.003	0.75	
FM (kg)			
Before	29.57 ± 4.41	27.08 ± 3.55	0.05 ^b
After	28.25 ± 4.45	26.87 ± 3.84	0.001 ^c
P value ^a	0.001	0.34	
FFM (kg)			
Before	46.86 ± 4.26	44.92 ± 2.73	0.08 ^b
After	47.57 ± 4.14	44.86 ± 2.93	0.22 ^c
P value ^a	0.004	0.83	
FM (%)			
Before	36.51 ± 5.54	34.24 ± 2.85	0.09 ^b
After	34.85 ± 5.38	33.89 ± 2.85	0.001 ^c
P value ^a	0.001	0.90	

Data are presented as mean ± SD. BMI; Body mass index, FM; Fat mass, FFM; Fat free mass, *; P value for paired t test, ^b; P value for Independent sample t test, and ^c; P value for ANCOVA: adjusted for total calorie intake, dietary vitamin E intake, age, physical activity and baseline values.

Angiogenic markers

The effects of vitamin E on angiogenic indices are shown in Table 4. The basal values of the angiogenic markers were not different between the two groups. In within-group analysis, VEGF, bFGF, Ang-1, and Ang-1/Ang-2 all had significant reductions in the vitamin E-supplemented group. In between-group comparisons, after adjustment for age, BMI, physical activity, total calorie intake, dietary vitamin E intake, and baseline values, supplementation with vitamin E had significant effects on VEGF, Ang-1, and Ang-1/Ang-2 ratio ($P = 0.01$, $P = 0.001$ and $P = 0.03$, respectively).

Table 4: Baseline and 8 weeks after intervention values of the serum angiogenic markers in the vitamin E and placebo groups

Variables	Vitamin E n=22	Placebo n=21	P value
VEGF (pg/mL)			
Before	733.15 (678.03, 1332.15)	423.40 (240.45, 1879.55)	0.96 ^b
After	329.85 (290.00, 1381.06)	420.00 (274.15, 1628.72)	0.01 ^c
P value ^a	0.005	0.48	
bFGF (pg/mL)			
Before	345.20 (305.99, 631.65)	370.10 (301.35, 590.45)	0.76 ^b
After	314.18 (231.95, 318.80)	386.00 (303.7, 642.75)	0.24 ^c
P value ^a	0.003	0.66	
Ang-1 (pg/mL)			
Before	1627.16 (1381.54, 2814.50)	1461.80 (1175.90, 1811.05)	0.28 ^b
After	864.80 (645.90, 1627.16)	1305.01 (1305.01, 1774.45)	0.001 ^c
P value ^a	0.001	0.87	
Ang-2 (pg/mL)			
Before	427.35 (247.78, 590.10)	432.49 (238.00, 493.45)	0.68 ^b
After	436.65 (250.70, 554.70)	410.91 (332.75, 410.91)	0.83 ^c
P value ^a	0.81	0.49	
Ang-1:Ang-2			
Before	3.44 (2.97, 5.27)	3.41 (2.72, 5.12)	0.49 ^b
After	2.63 (1.46, 3.74)	3.52 (3.17, 4.78)	0.03 ^c
P value ^a	0.03	0.61	

Ang-1; Angiopoietin-1, Ang-2; Angiopoietin-2, VEGF; Vascular endothelial growth factor, bFGF; Basic fibroblast growth factor, ^a; P value for Wilcoxon test, ^b; P value for Mann-Whitney U-test, and ^c; P value for ANCOVA: adjusted for total calorie intake, dietary vitamin E intake, age, physical activity and baseline values. Data are shown as median (25th, 75th).

Discussion

The present study was conducted to investigate the effect of vitamin E supplementation on the angiogenic markers in patients with PCOS. As far as we know, the present clinical trial is the first to examine the effects of vitamin E supplementation on serum angiogenic markers and anthropometric parameters in patients with PCOS. The results of this study revealed a significant reduction in weight and fat mass after eight weeks of supplementation with vitamin E among patients with PCOS. Both groups had lower energy intakes than daily estimated energy requirements (EER) for moderately active women. Low energy intake is considered a way of weight reduction, so, it is possible that the study subjects had reduced their calorie intakes for weight reduction. Only in the vitamin E group, weight reduction was significant. Few studies had assessed the effects of vitamin E supplementation on body composition components. There is some evidence about an inverse association between serum vitamin E concentration and adiposity (21). It was found that vitamin E is involved in the expression of some genes, like as leptin and peroxisome proliferator-activated receptor- γ (PPAR γ), which are related to the glucose and lipid metabolism (14, 22). Leptin regulates food intake and energy balance thus plays a key role in the regulation of body fat mass (14). PPAR γ is an adipogenic factor and acts as a regulator of adipogenesis (23). Increased PPAR γ activity may have a positive effect on body weight gain and FM (22). Vitamin E down-regulates the expression of PPAR γ (24).

Our study results indicated that vitamin E significantly lowered serum Ang-1 levels, while no change was observed in Ang-2 concentration in PCOS women. There is some evidence on angiopoietin disturbances in PCOS women. Scotti et al. (7) investigated angiopoietins of follicular fluids and reported an increase in Ang-1 but no changes in Ang-2.

In our literature review, there were no studies on vitamin E effects on the Ang-1 levels. The probable mechanism of reducing Ang-1 level by vitamin E may be linked with the reactive oxygen species (ROS). The increasing effects of ROS on the level, signaling and biological effects of Ang-1 were shown. Vitamin E has antioxidative properties, so, by scavenging of ROS, it decreases ROS and therefore, Ang-1 levels (25, 26).

In our study, the Ang-1/Ang-2 ratio was decreased. Restoration of the increased level of Ang-1/Ang-2 enhances vascular progression, which in turn, promotes proper follicular evolution and increased ovulation (27). The exact mechanism(s) by which vitamin E exerts these regulatory effects are still unknown, though some possible mechanisms have been proposed. It was stated that oxidants stimulate angiogenesis while antioxidants counteract angiogenesis (28). In addition, tocopherols exert their anti-angiogenic and anti-proliferative effects through preventing signaling and activation of PI3K/PDK/Akt signaling pathway, and inhibiting tube formation of endothelial cells (29).

Our study suggested a lowering effect for vitamin E intake on VEGF in PCOS women. There is some evidence indicating VEGF roles in the pathophysiology of PCOS (4, 30). VEGF, through neovascularization in the ovaries of PCOS patients, supports the increase in ovarian mass. Elevated levels of VEGF have been reported in women with PCOS (31). In addition, endocrine gland-VEGF, as an endothelial cell mitogen, has been shown to be over expressed in the PCOS patients' ovaries (32). Many studies assessed the effects of vitamin E on VEGF. These studies showed different effects for tocopherol on VEGF expression and angiogenesis (33, 34). It seems that the effects are dependent on the phosphorylation status of α -tocopherol (35). In an in vitro study, phosphorylated α -tocopherol (α TP) stimulated VEGF generation, while non-phosphorylated (α T) form did not (36). This is the outcome of PI3K/Akt signaling pathway stimulation or inhibition. Tocopherol phosphorylation and dephosphorylation may indirectly influence pro-angiogenic or anti-angiogenic activities. Creation of α TP in vivo probably describes pro-angiogenic effects of vitamin E. Placenta creation, inhibition of ischemia/reperfusion injury in the brain or cardiovascular system and promotion of wound healing are pro-angiogenic activities. Further, the pro-angiogenic ability of α TP is important in terms of expansion of solid tumors (37).

Our study did not demonstrate the effect of vitamin E on the bFGF in PCOS patients. In contrast to VEGF, little is known about agents influencing bFGF. bFGF has important functions in the ovarian angiogenesis. bFGF is a follicle-stimulating hormone, expressed in theca and granulosa cells leading to promotion of follicular growth and managing its activity (38). bFGF enhances angiogenesis by different mechanisms including stimulation of endothelial cell reproduction, chemotaxis and formation of matrix repairing enzymes such as plasminogen activator and collagenase (39). bFGF has been associated with obesity which is a common characteristic of PCOS. In addition, Artini et al. (40) reported higher levels of bFGF in serum and follicular fluids of patients with PCOS. Thus, correction of bFGF alterations in the biological fluids of PCOS women should be further examined.

Our study had some limitations. The most important limitation of the study was lack of measurement of vitamin E concentration at the baseline and at the end of study. In our study, the duration of the disease was not assessed. Another limitation was the small sample size. We could not provide a sonographic evaluation of ovarian masses at follow-up. Additionally, self-reported dietary intakes -which have the probability of under/over-reporting- and short duration of the intervention were other limitations for our study.

Conclusion

In patients with PCOS, vitamin E supplementation has useful effects on some anthropometric measurements

and Ang-1, VEGF and Ang-1/Ang-2 ratio in blood. These findings suggest possible beneficial effects for vitamin E on PCOS. Concerning our study limitations, further studies are recommended to explore the potential effects of vitamin E in the management of angiotensin disturbances among PCOS patients.

Acknowledgements

The present study was part of an M.Sc. thesis in nutrition and supported by a grant from Nutrition Research Center, and Student Research Committee, at the Vice-Chancellor for Research at Tabriz University of Medical Sciences (grant number: 5/D/4438). The authors would like to appreciate the support of Pourateb pharmaceutical company for providing vitamin E soft gels and Barij Esans Company, Kashan, Iran, for providing the placebo. The authors have no potential conflict of interest.

Authors' Contributions

S.S., Sh.T., A.I., M.P.; Participated in data collection, analysis of data, and drafting the manuscript. B.P.G.; Was involved in the theory and objective of the study, and preparation final revision of the manuscript. All authors read and approved the final version of the manuscript.

References

1. Goodman NF, Cobin RH, Futterweit W, Glueck JS, Legro RS, Carmina E. American Association of Clinical Endocrinologists, American College of Endocrinology, and Androgen Excess and PCOS Society disease state clinical review: guide to the best practices in the evaluation and treatment of polycystic ovary syndrome-part 1. *Endocr Pract.* 2015; 21(11): 1291-1300.
2. Setji TL, Brown AJ. Polycystic ovary syndrome: update on diagnosis and treatment. *Am J Med.* 2014; 127(10): 912-919.
3. Jalilian A, Kiani F, Sayehmiri F, Sayehmiri K, Khodae Z, Akbari M. Prevalence of polycystic ovary syndrome and its associated complications in Iranian women: a meta-analysis. *Iran J Reprod Med.* 2015; 13(10): 591-604.
4. Tal R, Seifer DB, Arici A. The emerging role of angiogenic factor dysregulation in the pathogenesis of polycystic ovarian syndrome. *Semin Reprod Med.* 2015; 33(3): 195-207.
5. Abramovich D, Irusta G, Bas D, Cataldi NI, Parborell F, Tesone M. Angiotensin/TIE2 system and VEGF are involved in ovarian function in a DHEA rat model of polycystic ovary syndrome. *Endocrinology.* 2012; 153(7): 3446-3456.
6. Berisha B, Schams D, Rodler D, Pfaffl MW. Angiogenesis in the ovary-the most important regulatory event for follicle and corpus luteum development and function in cow-an overview. *Anat Histol Embryol.* 2016; 45(2): 124-130.
7. Scotti L, Parborell F, Irusta G, De Zuñiga I, Bisioli C, Pettorossi H, et al. Platelet-derived growth factor BB and DD and angiotensin1 are altered in follicular fluid from polycystic ovary syndrome patients. *Mol Reprod Dev.* 2014; 81(8): 748-756.
8. Di Pietro M, Pascuali N, Parborell F, Abramovich D. Ovarian angiogenesis in polycystic ovary syndrome. *Reproduction (Cambridge, England).* 2018; 155(5): R199-R209.
9. Wells SR, Jennings MH, Rome C, Hadjivassiliou V, Papas KA, Alexander JS. alpha-, gamma-and delta-tocopherols reduce inflammatory angiogenesis in human microvascular endothelial cells. *J Nutr Biochem.* 2010; 21(7): 589-597.
10. Kurdoglu Z, Ozkol H, Tuluze Y, Koyuncu I. Oxidative status and its relation with insulin resistance in young non-obese women with polycystic ovary syndrome. *J Endocrinol Invest.* 2012; 35(3): 317-321.
11. Rahmani E, Samimi M, Afshar Ebrahimi F, Foroozanfar F, Ahmadi S, Rahimi M, et al. The effects of omega-3 fatty acids and vitamin E co-supplementation on gene expression of lipoprotein (a) and oxidized low-density lipoprotein, lipid profiles and biomarkers of

- oxidative stress in patients with polycystic ovary syndrome. *Mol Cell Endocrinol.* 2017; 439: 247-255.
12. Fatemi F, Mohammadzadeh A, Sadeghi MR, Akhondi MM, Mohammadmoradi S, Kamali K, et al. Role of vitamin E and D3 supplementation in intra-cytoplasmic sperm injection outcomes of women with polycystic ovarian syndrome: a double blinded randomized placebo-controlled trial. *Clin Nutr ESPEN.* 2017; 18: 23-30.
 13. Mondul AM, Rager HC, Kopp W, Virtamo J, Albanes D. Supplementation with alpha-tocopherol or beta-carotene reduces serum concentrations of vascular endothelial growth factor-D, but not -A or -C, in male smokers. *J Nutr.* 2011; 141(11): 2030-2034.
 14. Garcia OP, Long KZ, Rosado JL. Impact of micronutrient deficiencies on obesity. *Nutr Rev.* 2009; 67(10): 559-572.
 15. Lauritsen MP, Bentzen JG, Pinborg A, Loft A, Forman JL, Thuesen LL, et al. The prevalence of polycystic ovary syndrome in a normal population according to the Rotterdam criteria versus revised criteria including anti-Müllerian hormone. *Hum Reprod.* 2014; 29(4): 791-801.
 16. Izadi A, Ebrahimi S, Shirazi S, Taghizadeh S, Parizad M, Farzadi L, et al. Hormonal and metabolic effects of coenzyme Q10 and/or vitamin E in patients with polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2018; 104(2): 319-327.
 17. Samimi M, Zarezade Mehrizi M, Foroozanfard F, Akbari H, Jamilian M, Ahmadi S, et al. The effects of coenzyme Q10 supplementation on glucose metabolism and lipid profiles in women with polycystic ovary syndrome: a randomized, double-blind, placebo-controlled trial. *Clin Endocrinol.* 2017; 86(4): 560-566.
 18. Aghanejad Nozari R, Farshbaf-Khalili A, Sattarzadeh N, Asghari Jafarabad M. The effect of counseling on menstrual hygiene, physical activity, and nutritional status of female adolescent students: A randomized controlled field trial. *Crescent J Medical Biol Sci.* 2019; 6(3): 393-402.
 19. Estebarsari F, Shojaeizadeh D, Mostafaei D, Farahbakhsh M. Planning and evaluation of an educational program based on PRECEDE model to improve physical activity in female students. *Hayat.* 2010; 16(1): 48-54.
 20. Li H, Lei N, Zhang M, Li Y, Xiao H, Hao X. Pharmacokinetics of a long-lasting anti-VEGF fusion protein in rabbit. *Exp Eye Res.* 2011; 97(1): 154-159.
 21. Gunanti IR, Marks GC, Al-Mamun A, Long KZ. Low serum concentrations of carotenoids and vitamin E are associated with high adiposity in Mexican-American children. *J Nutr.* 2014; 144(4): 489-495.
 22. Joosen AM, Bakker AH, Zorenc AH, Kersten S, Schrauwen P, Westerterp KR. PPARgamma activity in subcutaneous abdominal fat tissue and fat mass gain during short-term overfeeding. *Int J Obes (Lond).* 2006; 30(2): 302-307.
 23. Liu GS, Chan EC, Higuchi M, Dusing GJ, Jiang F. Redox mechanisms in regulation of adipocyte differentiation: beyond a general stress response. *Cells.* 2012; 1(4): 976-993.
 24. Zhang Y, Li Y, Liang X, Gao J. Effects of dietary vitamin E supplementation on growth performance, fatty acid composition, lipid peroxidation and peroxisome proliferator-activated receptors (PPAR) expressions in juvenile blunt snout bream *Megalobrama amblycephala*. *Fish Physiol Biochem.* 2017; 43(4): 913-922.
 25. Harfouche R, Malak NA, Brandes RP, Karsan A, Irani K, Hussain SN. Roles of reactive oxygen species in angiopoietin-1/tie-2 receptor signaling. *FASEB J.* 2005; 19(12): 1728-1730.
 26. Prauchner CA. Angiogenesis inhibition by antioxidants. *International Journal of Biomedical Science and Engineering.* 2014; 2(6-1): 7-19.
 27. Di Pietro M, Parborell F, Irusta G, Pascuali N, Bas D, Bianchi MS, et al. Metformin regulates ovarian angiogenesis and follicular development in a female polycystic ovary syndrome rat model. *Endocrinology.* 2015; 156(4): 1453-1463.
 28. Kim YW, Byzova TV. Oxidative stress in angiogenesis and vascular disease. *Blood.* 2014; 123(5): 625-631.
 29. Saghiri MA, Asatourian A, Ershadifar S, Moghadam MM, Sheibani N. Vitamins and regulation of angiogenesis: [A, B1, B2, B3, B6, B9, B12, C, D, E, K]. *J Funct Foods.* 2017; 38: 180-196.
 30. Peitsidis P, Agrawal R. Role of vascular endothelial growth factor in women with PCO and PCOS: a systematic review. *Reprod Biomed Online.* 2010; 20(4): 444-452.
 31. El Behery MM, Diab AE, Mowafy H, Ebrahiem MA, Shehata AE. Effect of laparoscopic ovarian drilling on vascular endothelial growth factor and ovarian stromal blood flow using 3-dimensional power Doppler. *Int J Gynaecol Obstet.* 2011; 112(2): 119-121.
 32. Irani M, Seifer DB, Grazi RV, Irani S, Rosenwaks Z, Tal R. Vitamin D decreases serum VEGF correlating with clinical improvement in vitamin D-deficient women with PCOS: a randomized placebo-controlled trial. *Nutrients.* 2017; 9(4): 334.
 33. Chuang CH, Huang CS, Hu ML. Vitamin E and rutin synergistically inhibit expression of vascular endothelial growth factor through down-regulation of binding activity of activator protein-1 in human promyelocytic leukemia (HL-60) cells. *Chem Biol Interact.* 2010; 183(3): 434-441.
 34. Shibata A, Nakagawa K, Sookwong P, Tsuduki T, Oikawa S, Miyazawa T. delta-Tocotrienol suppresses VEGF induced angiogenesis whereas alpha-tocopherol does not. *J Agric Food Chem.* 2009; 57(18): 8696-8704.
 35. Zingg JM, Meydani M, Azzi A. alpha-Tocopheryl phosphate—An activated form of vitamin E important for angiogenesis and vasculogenesis? *Biofactors.* 2012; 38(1): 24-33.
 36. Zingg JM, Libinaki R, Lai CQ, Meydani M, Gianello R, Ogru E, et al. Modulation of gene expression by alpha-tocopherol and alpha-tocopheryl phosphate in THP-1 monocytes. *Free Radic Biol Med.* 2010; 49(12): 1989-2000.
 37. Zingg JM, Azzi A, Meydani M. Induction of VEGF expression by alpha-tocopherol and alpha-tocopheryl phosphate via PI3Kγ/PKB and hTAP1/SEC14L2-mediated lipid exchange. *J Cell Biochem.* 2015; 116(3): 398-407.
 38. Qiao J, Feng HL. Extra-and intra-ovarian factors in polycystic ovary syndrome: impact on oocyte maturation and embryo developmental competence. *Hum Reprod Update.* 2010; 17(1): 17-33.
 39. Tal R, Segars JH. The role of angiogenic factors in fibroid pathogenesis: potential implications for future therapy. *Hum Reprod Update.* 2014; 20(2): 194-216.
 40. Artini PG, Monti M, Matteucci C, Valentino V, Cristello F, Genazzani AR. Vascular endothelial growth factor and basic fibroblast growth factor in polycystic ovary syndrome during controlled ovarian hyperstimulation. *Gynecol Endocrinol.* 2006; 22(8): 465-470.