**Epigenetic Modifications, A New Approach to Male Infertility Etiology: A Review**

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**Abstract**

Recent studies have indicated that epigenetic alterations are critical for normal function and development of spermatocytes during the fertilization process. This review will focus on the latest advances in epigenome profiling of the chromatin modifications during sperm development, as well as the potential roles of epigenetic mechanisms in the context of male infertility. In this review, all data were collected from published studies that considered the effect of epigenetic abnormalities on human spermatogenesis, sperm parameters quality, fertilization process, embryo development and live births. The database PubMed was searched for all experimental and clinical studies using the Keywords “epigenetic modifications”, “male infertility”, “spermatogenesis”, “embryo development” and “reproductive function”. Post-translational modifications of histone, DNA methylations and chromatin remodeling are among the most common forms of epigenetic modifications that regulate all stages of spermatogenesis and fertilization process. Incorrect epigenetic modifications of certain genes involved in the spermatogenesis and sperm maturation may be a main reason of male reproductive disorder and infertility. Most importantly, abnormal patterns of epigenetic modifications or transgenerational phenotypes and miRNAs expression may be transmitted from one generation to the next through assisted reproductive techniques (ART) and cause an increased risk of birth defects, infertility and congenital anomalies in children. Epigenetic modifications must be considered as one of the main factors of unexplained male infertility etiology. Due to high risk of transmitting incorrect primary imprints to offspring, there is a need for more research into epigenetic alterations in couples who benefit of ART support.

**Keywords:** Epigenetics, Male Infertility, Reproductive Function, Sperm


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**Introduction**

Human infertility is now considered as a serious health problem affecting 15% of couples throughout the world (1). Male infertility factors contribute to about 50% of all infertility cases (2). It is a multifactorial syndrome that can be caused by numerous factors (Fig.1). Anatomical or structural defects such as varicocele and obstructive problems, ejaculatory failures, genital tract infections, hyperviscous semen, endocrinological disorders, impaired spermatogenesis, chromosomal abnormalities, and immunologic problems are the most common reasons for male infertility (3-5). Environmental or occupational factors such as smoking or opiate using, exposure to pesticides or radiation can influence gamete health and subsequently, male fertility (6). Males with normal sperm parameters, but unexplained cause are defined as idiopathic infertile men. Several factors such as oxidative stress caused by the massive production of reactive oxygen species (ROS), are categorized as the most common causes of this type of male infertility. It seems that ROS accompanied with sperm DNA fragmentation, gene mutations, and genetic disorders (7). The molecular genetic abnormalities rate in the male infertility etiology are notable, ~15-30%, that affect various physiological processes such as steroidogenesis, sperm production and development, maturation, function and quality. Knowing the molecular genetic factors, related genes and potential regulatory mechanisms may provide better understanding of this disease pathogenesis.

Chromosomal abnormalities, point mutations in single genes such as cystic fibrosis transmembrane receptor (CFTR) gene, microdeletions of the Y chromosome, mitochondrial genome (mtDNA) mutations, and imprinting disorders are the most important causes of the molecular genetic basis of infertility in men (8, 9). Also, mutations and polymorphisms in different genes,
which may adversely affect sperm production and male fertilization ability, can be associated with severe infertility in men (9). For instance, deletions of the Y chromosome, particularly azoospermia factor A, B and C regions (AZFa, AZFb and AZFc), are the most common causes of Y-linked male infertility which leads to severe damage in spermatogenesis (10). Additionally, deletions in various autosomal and X-linked genes such as androgen receptor (AR), and ubiquitin-specific protease-26 (USP26) genes, which are considered as critical genes of normal testis development and spermatogenesis, can be associated with male fertility disorder (11). Therefore, any changes such as translocations, inversions, mutations, deletions or insertion and etc. in the related chromosomes and/or genes can change the normal function of the male reproductive system and result in severe damage to male fertility.

Molecular basis of epigenetic alterations

The molecular mechanisms of epigenetic processes come from covalent modifications of protein and DNA components of chromatin. It consists of a range of modifications, including methylation of DNA, post-translational modifications of the core histone, and chromatin remodeling (17). These modifications are processed by three different enzymes, i.e. writer that adds epigenetic marks on DNA and histone, reader that recognizes epigenetic marks, and eraser responsible for removing these modifications (18).

Epigenetic regulations of gene expression by changing the chromatin structure and DNA accessibility. These modifications are extremely important for the development and differentiation of all cells, especially spermatozoa. Recent studies have illustrated that epigenetic modifications affect expression of various genes which are responsible for the development of the male reproductive system, sperm production, and male sexual behavior (19). This indicates that incorrect modifications of DNA can cause abnormal male sexual development and reproductive dysfunction (20). Epigenetic modifications not only control the process of spermatogenesis, but also they modulate the process of sperm maturation, sperm-egg interaction, fertilization, and embryo development. The interesting chemical changes of the DNA molecule, such as chromatin remodeling, methylation of DNA, histones post-translational modifications, and gene
DNA demethylation importance in the development replicated by DNMT1 (3). DNA demethylation is another DNA replication, while we encounter passive DNA methylation marks. Here, methylation occurs in a C5 position of the cytosine-phosphate-guanine (CpG) islands that have been located upstream of nearly 40% of mammalian genes. Here, methylation occurs in a C5 position of cytosine residues to create 5-methyl cytosine. In this process, the methyl group is originated from S-adenosyl methionine. Hypermethylation of DNA can suppress transcription by preventing transcription factors binding and inducing several transcriptional repressors binding (20). DNA methylation is controlled by several classes of DNA methyltransferases (DNMTs), including DNMT1, DNMT3a and DNMT3b, that serve as writers (21). All DNA methyltransferases serve as writers. DNMT enzymes regulate both initiation and maintenance of methylation marks.

DNA demethylation can occur through passive or active mechanisms. Active DNA demethylation occurs when a methyl group is removed independently during DNA replication, while we encounter passive DNA demethylation when newly synthesized DNA strands are replicated by DNMT1 (3). DNA demethylation is another process that prevents transmission of inappropriate epigenetic marks to the future generations. Despite of DNA demethylation importance in the development process, its action mechanism is unclear (22).

CpG regions methylation/demethylation, one of the important processes of gene expression regulation, affects transcription factors binding and then represses expression of imprinted and developmental genes, while its deficiency may result in some disorders (21). Correct DNA methylation plays critical role in the various essential cellular processes such as X- chromosome inactivation and chromatin stability (23).

Recent evidence has indicated that correct methylation of DNA is essential for fertility and embryo development. Also, abnormal DNA methylation pattern has been reported in spermatozoa of idiopathic infertile men with poor semen quality (24). Some studies demonstrated that there is a stable single CpG methylation pattern in sperm of fertile men, while in the subfertile men show significantly modified pattern (25). Interestingly, there is a correlation between DNA methylation level and sperm motility, sperm chromatin and DNA integrity (26, 27). For example, promoters’ hypermethylation of genes, including MTHFR, IGF2, H19, PLAG1, and SNRPN has been shown to be correlated with poor sperm quality and increased risk of infertility. Houshdaran et al. (28) found that hypermethylation of DNA, especially in the promoter regions of PLAG1, PAX8, DIRAS3, MEST, SFN, NTF3, and HRAS genes, can decrease counts, motility and normal morphology of sperm. Khazamipour et al. (29) reported hypermethylation of MTHFR promoter in nearly 53% of the men with non-obstructive azoospermia, while they observed no hypermethylation of MTHFR promoter in the men with obstructive azoospermia. Similarly, Wu et al. (30) found that the MTHFR gene promoter hypermethylation in human spermatozoa causes idiopathic infertility. These findings indicate that MTHFR hypermethylation is an important epigenetic abnormality which can contribute to male infertility. A more recent study has revealed that hypermethylation of SOX30 gene causes its silencing, impaired spermatogenesis and non-obstructive azoospermia in mice (31).

Abnormal methylation of the promoter regions of H19 and MEST genes can be associated with impaired spermatogenesis and oligozoospermia (32). Hammoud et al. (33), compared CpG methylation of LIT1, SNRPN, MEST, H19, PLAG1, PEG3, and IGF2 genes in spermatozoa of fertile and infertile men. Methylation of DNA in spermatozoa of infertile patients was significantly modified for all genes except IGF2. Furthermore, a significant relationship was observed between impaired spermatogenesis and hypermethylation of these six genes. Boissonnas et al. (34), found that H19 was considerably hypomethylated in men with teratozoospermia and oligoasthenoteratozoospermia. Another study indicated that MEST hypermethylation has more adverse effects on sperm quality than H19/IGF2-ICR1 hypomethylation. In this study, Rajender et al. (35) compared hypermethylation of MEST with hypomethylation of H19/IGF2.

The X-linked reproductive homeobox (RHOX) gene}

![Diagram](image.png)
clusters, are essential for normal spermatogenesis, germ cell survival and male fertility. These Homeobox genes are exclusively expressed in the reproductive system. A recent study has introduced ROXH cluster functions as an appropriate biomarker for idiopathic male infertility. This study also revealed that hypermethylation of ROXH cluster can cause severe abnormalities in the sperm parameters (36). It is also assumed that hypermethylation of DNA may suppress ROXH gene clusters transcription. Incorrect DNA methylation in the promoter region of the deleted-in-azoosperma-like (DAZL) gene has been shown to be associated with impaired spermatogenesis and sperm dysfunction (37). It is estimated that approximately 10% of men with impaired spermatogenesis suffered from DAZL deletion (38). Recently it has been reported that decreased expression of DNMTs genes is associated with azoospermia and infertility (39). Hammoud et al. (33), revealed a substantial relationship between modified methylation of H19, LIT1, SNRPN, MEST, PEG3, PLAG1, and IGF2 genes in human sperm and male infertility. In another study, DNA methylation at MEST gene was considerably correlated to increased level of FSH, decreased testicular volume and resulted in oligozoospermia (40). Also, it has been found that abnormal methylation reaches to about 41% in the sperm cells of patients who undergoing ART (41). Decreased level of DNA methylation in the H19-ICR has been reported that is negatively correlated with sperm count. A significant decrease of DNA methylation in the H19 gene was found in testicular sperm of azoospermic men in comparison with fertile men (42). In another study by Vieweg et al. (25), methylation of sperm DNA in normozoospermic men exhibited low standard deviation, while a higher variability in the percentage of DNA methylation was observed in the promoters of subfertile men. These data suggest that DNA methylation abnormalities in the human sperm impact on sperm parameters quality and male infertility.

**Histone modifications Role in male infertility**

Nucleosome, fundamental unit of chromatin, includes a short turn of DNA that wrapped around a histone octamer, two copies of each H2A, H2B, H3, and H4. This organization provides a rigid structure to chromatin. Histone modification is a covalent post-translational change on the lysine rich tail of any of the histone proteins, particularly H3 and H4 histones (43). Histone modifications serve as a key regulator for activation and inactivation of various genes. This reversible process is controlled by acetyltransferases, deacetyltransferases, methyltransferases and demethylases enzymes (44). Different types of post-transcription or modifications may occur by targeting the N-terminal tail of histones via acetylation, methylation, phosphorylating, ubiquitination, simulation, ADP-ribosylation, glycosylation, butyrylation, citrullination, and propionylation (Table 1) (45). However, histone acetylation, methylation and phosphorylation are the most frequently events during sperm development, spermatogenesis, fertilization and embryo development.

<table>
<thead>
<tr>
<th>Modification types</th>
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<tr>
<td>Acetylation</td>
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<td>Methylation</td>
<td>Lysine/Arginine</td>
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<tr>
<td>Phosphorylation</td>
<td>Serine/Threonine</td>
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<td>Ubiquitination</td>
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<td>Sumoylation</td>
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<td>Glycosylation</td>
<td>Serine/Threonine</td>
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<td>Butyrylation</td>
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<td>Propionylation</td>
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Histone modification is a key regulator of mitosis and spermatogenesis. Some studies have found that abnormal histone modifications during spermatogenesis may result in severe damage to sperm development and male fertility (35). The whole-genome of the male germ cells is entirely reorganized during the postmeiotic differentiation of spermatids into spermatozoa. While the core of the spermatid extends and condenses, most somatic histones are replaced progressively by protamines. Protamines are small basic sperm-specific nuclear proteins which are responsible for haploid genome condensation. This crucial process, tightly packaging, protects sperm genome against side effects of physical and chemical agents during fertilization process (46). The exact molecular mechanisms of histone replacement by protamines during the spermatogenesis process have been not well-understood. Recent evidence has indicated that epigenetic modifications of histones are probably the most important regulators for histones/protamines exchange in the human spermatozoa (47). However, histone/protamine exchange may differ in species because of the different amounts of protamine in mature mammalian sperm.

Histones acetylation, a reversible process, takes place at certain lysine residues on the amino-terminal ends of the core histones, which neutralizes the positive charge on the histones tails and therefore, decreases their affinity for DNA and provides open chromatin structure for transcription (48). In other hand, histone acetylation enhances the transcription level, such as acetylation of histones H3K4 (H3K4ac) and H3K9 (H3K9ac) (49). Histone acetyl transferases (HATs) and histone deacetylases (HDACs) are key regulators of histone acetylation/deacetylation (50). While HDACs suppress gene expression, HATs activity enhances gene expression. In other words, deacetylation is associated with gene expression suppression, but acetylation is associated with genes overexpression. Also, Histone acetylation throughout the human spermatogenesis has been reported. Increased level of histone H4 hyperacetylation (Hypac-H4) has been shown in spermatogonia, spermatocyte and also,
Histone H4 hyperacetylation facilitates interaction of transcription factors with the chromatin. Hypac-H4 in spermatids increases accumulation of bromodomain protein, which in turn triggers nuclear reorganization. Recently, Oikawa et al. (51), have revealed a significant decrease in the number of spermatogonia cells with Lys12ac-H4 or Hypac-H4. Several studies considered the relationship between Hypac-H4 and impaired spermatogenesis (47). A significant reduction in the number of spermatids with Hypac-H4 can be found in men with spermatogenesis deficiency and spermatid arrest. Shirakata et al. (44), demonstrated that hypac-H4 may trigger histones replacement by protamines during spermatogenesis. They also found different specific patterns of histone H4 modification during each stage spermatogenesis. For example, H4K8ac, H4K12ac, H4K5ac, H4K20me2 and H4me were increased in preleptotene spermatocytes, while H4me3 was decreased. These data suggest a significant association between histone H4 modifications and gene expression patterns during spermatogenesis process.

Recent evidence has identified several post-translational modifications, including S42 and K49 acetylation on protamine-1 (PRM1) and K64 acetylation on protamine-2 (PRM2). Although acetylation of histones is associated with genes transcription, sperm cells are transcriptionally inactive. Therefore, it can be hypothesized that histone acetylation displays an epigenetic mark which is transmitted from spermatozoa to oocyte. This event regulates expression of various genes involved in embryo development (25).

Histone methylation is another type of histone modifications which can be associated with activation or repression of chromatin state. For example, histone H3 trimethylation at lysine 36 (H3K36me3) and 4 (H3K4me3) are often associated with an open chromatin structure for active transcription, while H3 trimethylation at lysines 9 (H3K9me3) and 27 (H3K27me3) can cause transcriptional repression (25). A histone methyltransferases (HMTases) involved in methylation of histone that is generally associated with genes suppressing. Recent studies have shown different patterns of histone methylation in human spermatozoa. Therefore, abnormal methylation of histones can cause severe damages to spermatogenesis process and incorrect epigenetic programming in human spermatozoa (51).

Also, enzyme histone demethylases (HDMs), a member of the Jumonji protein family, involve in the histone demethylation. Recently, scientists have reported a tight link between the activity of JHMD2A (Jumonji C-terminal containing histone demethylase 2A) histone demethylase and spermatogenesis (47). JHMD2A regulates the expression of protamine 1 (Prm1) and transition nuclear protein 1 (Tnp1), which are necessary for condensation and proper packaging of sperm chromatin. A recent study has found histone H3 lysine 4 hypermethylation (H3K4me) or H3 lysine 27 (H3K27me) hypermethylation in sperm of infertile individuals compared to fertile subjects (25).

Histone phosphorylation, another kind of epigenetic modifications, is associated with transcription process activation and plays an important role in chromatin rearrangement during spermatogenesis (44). Also, Histone phosphorylation can regulate several biological events, including mitotic/meiotic chromosome condensation, activation and inactivation of genes transcription, and double-strand DNA breaks (DSB) repair (47). Phosphorylation of histones is also involved in regulation of meiotic replication, chromatin remodeling and compaction in the nucleus of spermatozoa. Therefore, defects in histone phosphorylation can be associated with sperm dysfunction and male infertility problems.

**Chromatin remodeling role in male infertility**

Chromatin remodeling is the dynamic rearrangement of chromatin structure in which several protein complexes such as a SWI/SNF, ISW1 and MI-2 proteins change the nucleosomes location and structure in an ATP-dependent process (52). During the chromatin remodeling process the condensed chromatin of spermatozoa transmits imprinting and epigenetic information to the embryo (53). Miller et al. (54), showed that incorrect DNA packaging in the mico spermatozoa causes infertility. Correct DNA packaging is essential for normal spermatogenesis because approximately 85% of the histones are substituted by protamines during the spermatogenesis process (55). During the initial stages of spermiogenesis, histones are hyperacetylated and undergo other epigenetic modifications. Nucleosomes are progressively disassembled in the final stage of spermatogenesis and then replaced by TNPs and eventually protamines.

Protamines incorporation into the chromatin of spermatozoa induces DNA condensation, which is critical for sperm production and migration. Phosphorylation/dephosphorylation of protamines is a key mechanism regulating interaction between them and DNA. Phosphorylation of protamines by a protein kinase A (PKA) increases their affinity or binding to DNA. Also, protamines dephosphorylation occurs concomitantly with the maturation of nucleoprotamine. Mutations in calcium/calmodulin-dependent protein kinase 4 (CaMK4), which is involved in phosphorylation of protamine 2, causes impaired spermiogenesis and consequently, male infertility (56). In the mature spermatozoa, DNA is packaged densely with protamines during the fertilization process, while the maternal genome is densely packaged with histones. Therefore, the highly condensed structure of nucleoprotamine must be unpacked and rearranged into a nucleosomal structure during the fertilization process. Incorrect epigenetic modifications during each stage of these processes can lead to severe male infertility. Given the critical roles of PRM1 and PRM2 in normal sperm function and fertilization process, the haploinsufficiency of these proteins can be associated with decreased amount of the respective protein, which in turn increases the...
percentage of sperm with abnormal chromatin structure and damaged DNA (35). Additionally, it has been shown that the optimal proportion of PRM1 to PRM2 (PRM1/PRM2 ratio), which is critical for normal fertilization, is tightly regulated (57). It has been indicated that the PRM1/PRM2 ratio in the fertile individuals ranges from 0.8-1.2 and deviation from this ratio may negatively affect sperm quality, sperm DNA integrity and increase the risk of infertility in men. Men with an increased PRM1/PRM2 ratio are characterized by poor quality of sperm and reduced fertilizing capacity. Aberrant H4 acetylation has been shown to be correlated to impaired spermatogenesis (42). Sonnack et al. (58), observed that a decreased level of histone H4 acetylation in men with infertility problems is significantly associated with impaired spermatogenesis. Histone H4 hyperacetylation, which decreases the affinity of histone to DNA, is essential for histone to protamine transition (35). Hyperacetylation of histone H4 is also found in the infertile men with Sertoli cell-only syndrome (SCOS) (59).

**Genetic imprinting role in male infertility**

Genetic imprinting is an epigenetic process in which the expression of alleles occurs in a parent-of-origin-specific manner. This process is a consequence of changes in methylation of CpG islands in one allele. Genetic imprinting determines the origin of genes expression from the parental or maternal genomes, a process which is critical for embryo development (41). There are multiple loci imprinting in human. For example, human spermatogonia from the fetal period are often unmethylated at H19 differentially methylated regions (DMR), while spermatogonia from the testis are highly methylated in DMR (35). Several imprinted genes such as the paternally imprinted GTL2 and H19 genes were previously considered in men with infertility problems (60). The relationship between reduced methylation of the paternal IGF2/H19 imprinting control region 1 (ICR1) and GTL2 imprints in spermatozoa of men with disturbed spermatogenesis indicates the importance of genetic imprinting during spermatogenesis. In a study, Poplinski et al. (61), demonstrated that fertile individuals have high levels of IGF2/H19-ICR1, while methylation of MEST shows a significant decrease. They also showed that hypermethylation of IGF2/H19-ICR1 and MEST has a correlation with low sperm counts. These data indicated that imprinting deficiencies in the IGF2/H19 ICR1 and MEST can be associated with idiopathic male infertility. Therefore, aberrant MEST methylation can be considered as a strong indicator of sperm quality. Recent evidence has indicated that ART techniques such as intracytoplasmic sperm injection (ICSI), in vitro fertilization (IVF) and round spermatid injection (ROSI) can enhance the prevalence of imprinting defects and also, negatively affect human embryonic development through selecting immature sperm cells which may not have proper imprints or global methylation (3). More importantly, ART techniques may transmit epigenetic modifications to the next generation and cause the birth of children with a higher risk of infertility and other congenital abnormalities. Therefore, more efforts are necessary to optimize current ART techniques. Additionally, selection of appropriate procedures needs careful evaluation. Since epigenetic modifications can be maintained throughout the human lifespan and can potentially be transmitted to the next generations, long-term follow-up and health evaluation of ART offspring are essential to provide more robust clinical evidence (62).

**miRNAs role in the male infertility**

Epigenetic modifications may also extend on mechanisms, including non-coding RNA such as micro-RNAs (miRNAs). Non-coding RNAs, especially miRNAs and PIWI interacting RNAs (piRNAs), consist the most contents of RNAs in a cell and regulate gene expression at different levels. Also, miRNAs negatively regulate the expression pattern of various genes at the post-transcriptional level. Recent investigations confirmed the role of miRNAs in the regulation of development, differentiation, and progression of various diseases. Expression pattern alterations of these RNAs in the sperm cells can be associated with severe abnormalities of these cells and next generations (12). In an experimental study, Skinner et al. (63), demonstrated that alteration in the DNA methylation and ncRNA of sperm by Dichloro Diphenyl Trichloroethane (DDT) can be transferred and inherited in the next generation.

**Epigenetic regulation of sperm development**

Epigenetic modifications control all steps of fertilization from testicular cells development to embryo development (Fig.2). The initial stage of reprogramming event takes place in the developing gonads. The testes of all mammals involve all stages of spermatogenesis from germ cells to mature spermatozoa. In the normal germ cells, epigenetic alterations are reversible and cause changes in the various genes expression whenever it is necessary. Epigenetic modifications are highly active in developing germ cells, where sperm and oocyte are equipped with an appropriate epigenetic information before embryonic development (15). Germ cells undergo a series of dramatic morphological alterations, as well as specific and extensive chromatin remodeling during their differentiation into mature spermatozoa (64).

Spermatozoa are highly specialized cells, which are produced during spermatogenesis from germ cells. It is a highly conserved process that regulates by extensive cellular, epigenetic and chromatin modifications. Sperm cells transfer the genetic information from one generation to the next and are thus critical for maintenance of life. Genetic and epigenetic mechanisms, especially unique gene expression patterns, histone modifications and DNA methylation, have important functions in the regulation of germ-cell development (45). Meiosis, a process in which a germ cell is divided to four gametes, is associated with
histone modifications. After meiosis division, germ cells undergo several chromatin rearrangements, especially histone replacement with protamine.

Dynamic epigenetic alteration is central for differentiation of mammalian sperm; however, the mechanism of these modifications is still unknown. Spermatogenesis is a unique process which is precisely regulated by epigenetic modifications and results in the formation of haploid spermatozoa with the fertilization ability. The process of spermatogenesis is accompanied by chromatin rearrangement from a nucleosomal histones-based structure to a protamines-based structure. Mitotic amplification, meiotic, and post-meiotic phase (also known as spermiogenesis) are the critical phases of spermatogenesis. Replication and differentiation of spermatogonial stem cells into primary spermatocytes is the initial phase of spermatogenesis process. In meiosis, the primary spermatocytes are developed into haploid secondary spermatocytes through genetic recombination. During the haploid stage of spermatogenesis, round spermatids are differentiated into spermatozoa (65). In this process, histone H4 hyperacetylation causes nucleosome disassembly, DNA breaks and incorporation of non-allelic variants of histones (43). Histones are largely exchanged by transition proteins and then by the protamines PRM1 and PRM2. Recent evidence has indicated that ~99% of histones in the mice mature spermatozoa and ~85% of histones in the human sperm are exchanged by PRM1 and PRM2. Human sperm protamines are phosphorylated at several specific sites, including PRM1S11, PRM2S59 and PRM1S9; however, the exact mechanism of this phosphorylation is still unclear. These proteins also undergo further post-translational modifications during sperm maturation and transition in the epididymis. During this process, protamines form disulfide bonds and zinc bridges which stabilize the sperm chromatin structure (57). The relationship between protamines and DNA results in significant structural and molecular remodeling which subsequently compacts sperm DNA in the nucleoprotamine toroidal structures. The protamine-DNA interaction not only facilitates the normal morphology and hydrodynamic shape of the sperm head, but also protects the paternal genome against physical and chemical damages.

During the fertilization process, oocyte and spermatozoa fuse to form a single totipotent cell called zygote which can produce a new organism. Although sperm chromatin is extensively rearranged, histones are associated with the paternal genome postfertilization process and contribute to zygotic chromatin. The highly compacted sperm DNA is decondensed from its transcriptionally inactive state and expands to active state found in the paternal pronucleus. In the zygote, the paternal histone proteins might then serve as a template for newly synthesized histones during replication. In postfertilization, the programmatic sperm DNA packaging can therefore deliver epigenetic information to the oocyte and the zygote (53).

The patterns of DNA methylation are first acquired during gametogenesis. A growing number of evidences has illustrated the importance of DNA methylation for male germ cell development in a mouse model. It has been shown that gene targeting of enzymes responsible for DNA methylation leads to loss of germ cells methylation, meiosis deficiency and male infertility (42). A recent study has demonstrated that mono-, di- and tri-methylations of different histones, H3K9, H3K4, and H3K27, tightly regulate temporal expression and correct progression of spermatogenesis (44). Methylation level of H3K4 is enhanced in the spermatogonial stem cell. Reduced methylation level of histone H3K4 decreases the activity of Mll2 (an H3K4 methyl transferase) which in turn declines the number of spermatoocytes. This suggests the important role of H3K4 methylation in differentiation of spermatocyte from the stem cell stage (64). Conversely, the methylation of histones H3K9 and H3K27 in the stem cell is low, but its level increases during meiosis. Several lines of studies indicated the association between the methylation of histone H3K9 and euchromatin, heterochromatin and sex chromosomes in the late pachytene stage. However, the H3K9 methylation level decreases during the completion of meiosis, which is associated with an increase in H3K4me levels.

**Conclusion**

The advent of new technologies has revealed differential expression patterns of various genes in testicular cells and spermatozoa which opened a new window to male infertility etiology. Epigenetics alterations of DNA, histones, chromatin and miRNAs are significant factors that regulate spermatogenesis and impact fertility. These modifications not only involve in the regulation of somatic and germ cells development, but also control all stages of spermatogenesis process. Epigenetic alterations also control the correct expression patterns of various genes during the spermatogenesis and subsequently, fertilization. In other words, epigenetic alterations are necessary for the correct expression of various genes in normal conditions. Although ART technologies such as ICSI and IVF have provided a good opportunity for most infertile couples to have their own child, the abnormal epigenetic information transmission to the next generation may associate with increased risk of infertility and other health problems.

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**Authors’ Contributions**

A.B.H., I.L., H.Sh, H.I.; Were contributed to data collection and evaluation, drafting and statistical analysis, and manuscript writing. E.T.M.; Was contributed to manuscript writing, study concept and design, drafted or provided critical revision of the manuscript. A.Sh.; Was responsible for overall supervision. All authors read and
References


